The use of *in vitro* human biomarkers from relevant primary cell systems, to assess the effects of experimental and commercial e-liquids

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1. Introduction and Objectives

1.0 Introduction

As part of the ongoing stewardship of electronic cigarettes, and in line with the National Research Council "Toxicity Testing in 21st Century: A Vision and a Strategy" and subsequent documents, Fontem Ventures B.V., have investigated the utility of a new technique offered by DiscoverX. DiscoverX, is a leading supplier of in-house cell-based assays and services for the drug discovery and development industries. The BioMAP[®], Diversity PLUS product was chosen, consisting of 12 primary human cell-based systems from multiple tissues designed to model different aspects of the human body in an *in vitro* format (see Table 1). Cells are cultured either alone or as co-cultures and stimulated with a combination of biological proprietary factors (e.g. cytokines, growth factors, mediators, etc.) to recreate the multi-component signalling networks seen in normal cells. The identification of and disruption of key cellular signalling pathways are one of the main focuses of the NRC report. The Diversity PLUS panel consists of 148 biomarker readouts and has been used as a tool for drug discovery, competitive analysis and as a comparison to clinical standards of care. Each test agent generates a signature BioMAP [®] profile that is created from the changes in protein biomarker readouts within individual system environments. Each readout is then measured quantitatively by immune-based methods that detect proteins released into the cell media (e.g., ELISA) or functional assays that measure proliferation and cell viability.

1.1 Objectives

In the first instance, base e-liquids (without flavours) with and without nicotine were used to determine the suitability of the BioMap[®] system. Next subsequent testing evaluated the effects of commercial flavoured e-liquids on the system. Additionally, the osmolality of the diluted e-liquids was also measured to examine the potential effects of osmotic stress due to high concentrations of Propylene glycol (PG) and Vegetable glycerine (VG) being added to the systems (Iskandar et al., 2016; Gonzalez-Suarez et al., 2017).

2. Overview of BioMap ® Plus Panel

2.0 BioMap[®] Plus panel

Various disease states are potentially captured by the panel of 12 human primary cell lines. Vascular biology is modelled in both a Th1 (3C system) and a Th2 (4H system) inflammatory environment, as well as in a Th1 inflammatory state specific to arterial smooth muscle cells (CASM3C system). Additional systems recreate entire aspects of the systemic immune response including monocyte-driven Th1 inflammation (LPS system) or T cell stimulation (SAg system), chronic Th1 inflammation driven by macrophage activation (**/Mphg system**) and the T cell-dependent activation of B cells that occurs in germinal centres (BT system).

The BE3C system (Th1) and the BF4T system (Th2) represent potential markers of airway inflammation of the lung, whilst the **MyoF system** models myofibroblast-lung tissue remodelling. Lastly, skin biology is addressed in the KF3CT system modelling Th1 cutaneous inflammation and the HDF3CGF system models potential wound healing. Whilst disease progression is not currently well understood its entirety, inflammation is one of the key process that has been implicated in a number of diseases.

Table 1: Systems in BioMAP PLUS panel							
System Name	lcon	Cell	Disease				
3C	8	Venular endothelial cells	Cardiovascular Disease, Chronic Inflammation				
4H	8	Venular endothelial cells	Allergy, Asthma, Autoimmunity				
LPS		Peripheral blood mononuclear cells, Venular endothelial cells	Cardiovascular Disease, Chronic Inflammation				
SAg		Peripheral blood mononuclear cells, Venular endothelial cells	Autoimmune Disease, Chronic Inflammation				
ВТ	80	B cells, Peripheral blood mononuclear cells	Allergy, Asthma, Autoimmunity, Oncology				
BF4T	••••	Bronchial epithelial cells, Dermal fibroblasts	Allergy, Asthma, Fibrosis, Lung Inflammation				
BE3C		Bronchial epithelial cells	COPD, Lung Inflammation				
CASM3C	9	Coronary artery smooth muscle cells	Cardiovascular Inflammation, Restenosis				
HDF3CGF	-	Dermal fibroblasts	Chronic Inflammation, Fibrosis				
KF3CT		Dermal fibroblasts, Keratinocytes	Dermatitis, Psoriasis				
MyoF		Lung fibroblasts	Chronic Inflammation, Fibrosis, Matrix Remodeling, Wound Healing				
/Mphg		Macrophages, Venular endothelial cells	Cardiovascular Disease, Chronic Inflammation, Restenosis				

References

- Gonzalez-Suarez et al., (2017) App In vitro Toxicol 3(1): 41-55.

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3. Results: Effects of increasing concentrations of nicotine in base e-liquids and commercial e-liquids on the BioMAP® profile





- Some limitations of the assay include a reduced metabolic capacity of the cells and no liver model.
- The use of human primary cells are highly relevant model for the assessment of potential effects in humans.

Future work

- To use e-vapour aerosol extracts in the assay and use of a comparator product (i.e. cigarette) to see relative changes.
- To refine the exposures to physiologically relevant concentrations.

NRC (2007) Toxicity testing in 21st Century- A vision and a strategy <u>https://www.nap.edu/catalog/11970/toxicity-testing-in-the-21st-century-a-vision-and-a</u>: Iskander et al., (2016) Toxicol Mech Methods 26(6) 389-413.

BF4T	BE3C v	CASM3C	HDF3CGF	KF3CT	MyoF
	tPA	IL6 SAA M-CSF			
					Profiles FTM Com Com
CXCL8/1-80 IL-1alpha MMP-3 MMP-3 MMP-3 FAL-1 SRB FAL-1 FAL-1 FAL-1 LPA	CC54/ICAM-1 CC54/ICAM-1 CCXCL10/IP-10 CCXCL3/MIG CCXCL3/MIG CCXCL3/MIG CCXCL3/MIG CCXCL3/MIG EGFR HLA-DR HLA-DR MMP-3 RMP-3 SRB SRB SRB	CCL2/MCP-1 CD106/VCAM-1 CD106/VCAM-1 CD142/TIrsue Factor CD87AJPAR CCR37AJPAR CCC13/MIG HLA-DR HLA-DR HLA-DR IL-6 LDLR M-CSF PAL1 Proliferation Serum Amyloid A SRB	CCL2/MCP-1 CD106/NCAM-1 CCB4/ICAM-1 CCB1agen III CCL10/IP-10 CXCL9/MIG CXCL9/MIG EGFR MMP-1 PAL1 PAL1 PAL1 PAL1 PAL1 PAL1 PAL1 PAL	CCL2/MCP-1 CC54/ICAM-1 CCCL10/IP-10 CCCL13/MIG CCCL13/MIG IL-1alpha MMP-3 CCCL3/MIG IL-1alpha MMP-3 SRB SRB	alpta-SM Actin bFGF CD106/VCAM-1 Cd1agen 11 Cd1agen 12 Cd1agen 12

4. Conclusions

The addition of nicotine to base liquid gave rise to a significant decrease in biomarkers which were concentration related, in addition to those observed by PG/VG base liquid itself. Certain cell lines relating to BT (peripheral blood mononuclear cells), BEC3 (Bronchial epithelial cells) and HDF3CGF/KF3CT (dermal fibroblasts) and LPS peripheral blood cells and endothelial cells were most affected. The effects of adding a commercial flavour to the base liquid appear to be minor compared to the effects of adding nicotine

The assay appears to potentially be a useful tool for both comparisons between products and adds significantly to a weight of evidence risk assessment approach.



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Chart 1: Results of a base e-liquid (BL 1 containing only 50:50 PG and VG); **BL 2** containing 2.4% nicotine and BL 3 with a nicotine content of 4.5% were tested concentrations ranging between 0.031 to 4% added directly to media. The grey area in the middle of the chart represents the (95% historical control values confidence The interval). Y-axis represents a log-transformed ratio of the biomarker readouts for the test agent-treated sample (n = 1) over vehicle controls (average of \geq 6 vehicle controls from the same plate), cell variability, with values outside of this grey area (positive or negative) being significant.



Chart 2: There is an effect of adding commercial e-liquid A above that of the base liquid BL1 in the cell lines LPS, Sag, BT, Mphg at 0.5% with a considerably more marked effect of adding nicotine at 4.5% w/w to the commercial e-liquid B in the following cell lines 3C, LPS, Sag; BT; HDF3CGF. Concentrations were chosen to be neither cytotoxic or anti-proliferative. At both of the concentrations used osmolality was not considered to be an issue being in the range of 300-400 milliosmoles/litre.